Trace metal distribution in *Tilapia zillii* and *Synodontis schall* from the Siluko River, Edo State, Nigeria

Egun Nkonyeasua Kingsley^{1*},

Olowo Ukpamufo Cyril¹,

Biose Ekene²,

Jacob N. Jacob³,

Oboh Ijeoma Patience¹

¹ TETFund Centre of Excellence in Aquaculture and Food Technology, University of Benin, Benin City, Nigeria

² Department of Environmental Management and Toxicology, University of Benin, Benin City, Nigeria

³ Environmental and Analytical Research Laboratory, Department of Chemistry, University of Benin, Benin City, Nigeria Metal pollution of aquatic ecosystems has raised concerns about their uptake by aquatic organisms and subsequent biomagnification along the aquatic food chain. This study determined trace metal distribution in Tilapia zillii and Synodontis schall from the Siluko River in Edo State, Nigeria. Fresh fish samples were harvested monthly from September 2021 to August 2022 with the assistance of artisanal fishermen. Trace metal contents in the gills, liver, and muscles were determined using atomic absorption spectrophotometer. The metal pollution index (MPI) was used to estimate the extent of metal bioaccumulation in the fishes. Results showed species variation in metal accumulation in various organs: T. zillii had a higher mean content of Cu, Zn, Mn and Fe in all the organs, while S. schall had a higher mean content of Ni, Cr, and Pb in all the organs. The MPI values ranged from 5.28 to 5.94 in the organs of T. zillii and S. schall. The order of MPI values in *T. zillii* was muscles > liver > gills; in *S. schall*, it was liver > gills > muscles. The results indicate trace metal contamination of fishes due to aquatic pollution. High MPI values is an indication of the degree of metal pollution of the Siluko River and the high susceptibility of these metals to bioaccumulation in the fishes. This predisposes the fishes to metal toxicity and portends health risk to fish consumers. The continuous monitoring of activities within the river watershed to mitigate heavy metal pollution is recommended.

Keywords: aquatic toxicity, bioaccumulation, metal toxicity, metal pollution index (MPI), Siluko River

INTRODUCTION

The increasing exposure of freshwater bodies to the influx of diverse forms of pollutants from various anthropogenic activities within their water-

 * Corresponding author. Email: kingsley.egun@uniben.edu

shed has resulted in the deterioration of their water quality and raised concerns about the sustainability of the aquatic ecosystem for optimal production (Egun, Oboh, 2022a; Egun, Oboh, 2023; Biose et al., 2024). Trace metals are elements that are present in small amounts in the environment and in living organisms. Such trace metals as iron, zinc,

copper and manganese are essential elements for nutrition and physiology, as their deficiencies may result in organism malfunctioning; other trace metals, such as nickel, chromium and lead, have no known biological function and cause toxic effects and portend health risk to organisms in the environment even at low concentrations (Rainbow, 2007). The aquatic ecosystem is the final receptor of the released harmful substances, leading to the uptake and accumulation of these contaminants in the aquatic organisms. Trace metal contamination of aquatic ecosystems is an issue of global concern (Islam et al., 2020; Zahoor et al., 2023; Tesfaye et al., 2024), and particularly in the developing countries such as Nigeria (Enuneku et al., 2013; Egun, Ogiesoba-Eguakun, 2018; Egun, Oboh, 2022a; Egun, Oboh, 2023; Biose et al., 2024) as it poses a threat to achieving the United Nations sustainable development goals of life below water (Goal 14), provision of clean water (Goal 6) and good health and wellbeing (Goal 3).

The trophic position of fishes in the aquatic food web and their ability to accumulate metals from various sources, including sediments and water, identify them as bioindicators of trace metal contamination in aquatic ecosystems (Tesfaye et al., 2014; Muhammad, Ahmad, 2020). Also, the accumulation of trace metals in various tissues of fishes gives credence to exposure to polluted aquatic environment and could be used to evaluate the health condition of the environment from which they were collected (Qadir, Malik, 2011). Increasing global demand for fish meat as source of animal protein and the contribution of capture fisheries to household nutritional needs (Egun et al., 2024) emphasize the importance of assessing trace metal levels in fish populations in the wild (Bawuro et al., 2018; Chen et al., 2022; Muneer et al., 2022). The increase or decrease in the levels of certain pollutants in water bodies have been reported to directly or indirectly affect the behavioural and physiology of fishes (Egun, 2021). Reported studies on trace metal pollution of aquatic ecosystems globally (Custer et al., 2000; Juśkiewicz, Gierszewski, 2002; Basooma et al., 2021; Tesfaye et al., 2024), and freshwater

bodies in Nigeria (Oboh, Agbala, 2017; Egun, Oboh, 2022a; Egun, Oboh, 2023; Biose et al., 2024) have raised concerns about their uptake by aquatic organisms and subsequent biomagnification along the aquatic food chain (Egun et al., 2023; Uwaifo et al., 2023; Egun et al., 2025).

The fish species – *Tilapia zillii* and *Synodontis schall* – are commercial freshwater fish species, with high culinary acceptability in Nigeria. Their omnivorous feeding habit and transitional position in the trophic niche between primary producers and piscivores in the aquatic ecosystems has made them valuable bio-indicators of aquatic pollution and biomagnification of contaminants. Therefore, the aim of this study was to evaluate the distribution of trace metal concentrations in the organs of *T. zillii* and *S. schall* from the Siluko River as an indicator of aquatic biota contamination and a potential source of toxic trace metals (TTEs) to consumers.

MATERIALS AND METHODS

Study location

This study was carried out at the Siluko River (latitudes 06°32'22.5" N and 06°31'59.7" N and longitudes 005°09'32.5" E and 005°09'20.8" E), in Edo State, Nigeria (Figure). The Siluko River receives water from the Owena River and drains into the Atlantic via the Benin River (Oboh, Agbala, 2017). The river is flanked by secondary vegetation of rubber trees (*Hevea brasiliensis*), Awolowo weed (*Chrommolaena odorata*), African white mahogany, (*Khaya anthotheca*), Itako (*Stromboisa pustulata*), palm oil (*Elaeis guinensis*), bamboo tree (*Bambusa* spp.), and shrubs. The river is also characterised by floating vegetation such as duck weed (*Lemna* spp.) and water hyacinth (*Eichorrnia crassipes*).

Sample collection

Fresh samples of *T. zillii* (average weight 480 g) and *S. schall* (avg. wt. 430 g) were harvested monthly for a duration of 12 months (September 2021 to August 2022) using fishing nets and local fishing traps with the assistance of artisanal fishermen, and subsequently identified using taxonomic guides of Idodo-Umeh (2003). Fish

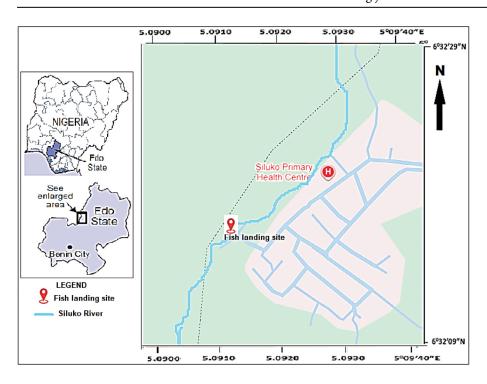


Figure. Map of Siluko River with fish landing site in Siluko Town, Edo State. Insert maps: (A) Nigeria (B) Edo State

samples were collected from fish catches intended for sale in the local fish market to consumers and fish processors. Fish samples were packed in polyethylene bags and preserved in ice boxes. The analysis of heavy metal contents in the fish species was conducted at the national reference laboratory – the Benin Owena River Basin Authority/University of Benin Analytical laboratory.

Laboratory analysis

In the laboratory, fresh fish samples were dissected and gutted, and tissues of interest (muscles, gills, and liver) were extracted. The extracted sample tissues were wrapped in aluminium foil paper and oven -dried at a temperature of 105°C for 1 h. Sample preparation and digestion using nitric acid (HNO₃) and perchloric acid (HClO₄) were carried out following the procedure described by APHA (2012). Trace metal concentrations in the sample filtrates were determined using the flame atomic absorption spectrophotometer (FAAS) (Model 210 VGP, Buck Scientific).

Quality assurance and control

Analysis was done in triplicates for all samples with mean values reported. Trace metal content in sample filtrates for the respective tissues was analysed using the FAAS, which was calibrated using Buck-certified atomic absorption standards for several trace metals to obtain a calibration curve. Details of certified reference materials, detection limits, quantification limits, and percentage material recovery on each metal are presented in Table 1.

To preserve the biological integrity of tissues of interest, collected fish samples were immediately placed in polyethylene bags and kept in an ice box which was used to carry the samples to the laboratory. A national reference laboratory – the Benin Owena River Basin Authority/ University of Benin Analytical laboratory, University of Benin, Nigeria was utilized for laboratory analysis.

Metal pollution index (MPI)

The total amount of metals found in the fish samples was compared using the metal pollution index (MPI) as described by USEPA (2011). The MPI was determined for each examined organ using the equation:

$$MPI = (M_1 \times M_2 \times M_3 \times M_4 \times M_5 \times M_6 \times M_7)^{1/n}$$

where *M* is the concentration of metals measured and expressed (mg/kg), and *n* is the number of metals measured.

Table 1. Certified values for elements in SRM 3233 detection limit (DL), quantification limit (QL), % of relative standard deviation (RSD) and recovery

Metals	Detection limits (DL) (μg mL ⁻¹)	Quantification limit (QL) (µg mL ⁻¹)	Relative standard deviation (%RSD)	% recovery
Fe	0.1-1.0	0.01-1.0	1–5	99
Zn	0.1-1.0	0.01-1.0	1–5	99
Cu	0.1-1.0	0.01-1.0	1–5	90
Mn	0.1-1.0	0.01-1.0	1–5	90
Ni	0.1-1.0	0.01-1.0	1–5	90
Cr	0.1-1.0	0.01-1.0	1–5	99
Pb	0.1-1.0	0.01-1.0	1-5	99

Source: Standard Reference Material (SRM) 3233 (2020).

Data analysis

All statistical analysis was computed using Microsoft Excel and Statistical Package for Social Sciences (SPSS) v.21. At every sampling visit, five specimens of each species were collected from an artisanal fish landing site in Siluko community. Analytical results are presented as the pooled means \pm SD per kg of each fish species, and ANOVA (p < 0.05) was used to de-

termine the variation in the trace metal content among the selected organs in fish species.

RESULTS

The mean concentrations of metals (Fe, Zn, Cu, Mn, Ni, Cr, and Pb) in gills, liver and muscles of *T. zillii* and *S. schall* from the Siluko River are presented in Table 2. The analytical results

Table 2. Summary of the metal content in the organs of T. zillii and S. schall from the Siluko River

Metal (mg/kg) of		Gills	Liver	Muscles	p value
wet weight		Mean ± SD	Mean ± SD	Mean ± SD	
Iron (Fe)	T. zillii	74.56 ± 1.08^{a}	84.84 ± 1.38^{b}	74.51 ± 0.35^{a}	<i>p</i> < 0.05
	S. chall	68.50 ± 6.16^{a}	62.5 ± 3.04^{a}	61.40 ± 7.23^{a}	<i>p</i> > 0.05
Zinc (Zn)	T. zillii	15.02 ±0.05 ^a	21.90 ± 0.25^{b}	17.60 ± 0.42^{b}	<i>p</i> < 0.05
	S. schall	13.67 ± 1.26^{a}	17.96 ± 0.70^{b}	11.66 ± 1.52^{a}	<i>p</i> < 0.05
Copper (Cu)	T. zillii	12.25 ± 0.39^{a}	10.89 ± 0.54^{a}	17.43 ± 0.70^{b}	<i>p</i> < 0.05
	S. schall	9.65 ± 0.51^{a}	9.25 ± 2.02^{a}	16.01 ± 1.25^{b}	p < 0.05
Manganese (Mn)	T. zillii	6.55 ± 0.39^{a}	8.32 ± 0.89^{a}	6.73 ± 0.30^{a}	<i>p</i> > 0.05
	S. schall	5.87 ± 0.61^{a}	6.02 ± 0.10^{a}	5.41 ± 0.48^{a}	<i>p</i> > 0.05
Nickel (Ni)	T. zillii	4.85 ± 0.19^{a}	4.20 ± 0.34^{a}	4.91 ± 0.06^{a}	<i>p</i> > 0.05
	S. schall	5.56 ± 0.07^{a}	5.25 ± 0.97^{a}	5.74 ± 0.86^{a}	<i>p</i> > 0.05
Chromium (Cr)	T. zillii	6.99 ± 0.21^{a}	4.91 ± 0.57^{a}	6.91 ± 0.08^{a}	<i>p</i> > 0.05
	S. schall	7.85 ± 0.38^{a}	7.89 ± 0.34^{a}	8.04 ± 0.72^{a}	<i>p</i> > 0.05
Lead (Pb)	T. zillii	0.04 ± 0.002^{a}	0.06 ± 0.004^{a}	0.05 ± 0.006^{a}	<i>p</i> > 0.05
	S. schall	0.05 ± 0.005^{a}	0.05 ± 0.008^{a}	0.04 ± 0.006^{a}	<i>p</i> > 0.05

Note: values are presented as pooled means \pm standard deviations (SD) of the metal content in the organs of fish species throughout the duration of the study. Across each row, similar superscripts indicate no significant difference (p > 0.05), while dissimilar superscripts indicate significant difference (p < 0.05).

showed that *T. zillii* had a higher mean content of Fe, Zn, Cu and Mn, while *S. schall* had a higher mean content of Ni, Cr, and Pb in all the organs. In both fish species, the content of Fe was the highest and that of Pb the lowest across the organs. The order of heavy metal contents in the gills of *T. zillii* and *S. schall* was Fe > Zn > Cu > Cr > Mn > Ni > Pb. For liver, the order was Fe > Zn > Cu > Cu > Cr > Mn > Ni > Pb in *T. zillii* and Fe > Zn > Cu > Cr > Mn > Ni > Pb in *S. schall*. In the muscles, the order was Fe > Zn > Cu > Cr > Mn > Pb in *T. zillii* and Fe > Cu > Cr > Ni > Pb in *T. zillii* and Fe > Cu > Cr > Ni > Pb in *S. schall*.

Metal pollution index (MPI)

The MPI values for the examined organs in *T. zillii* and *S. schall* varied from 5.28 to 5.94 (Table 3). The order of MPI values in *T. zillii* was muscles > liver > gills; in *S. schall*, it was liver > gills > muscles. *T. zillii* recorded higher non-significant MPI values in all the organs examined of both fish species.

Table 3. Metal pollution index (MPI) of organs of *T. zillii* and *S. schall* from the Siluko River

Ougana	MPI (mg/kg)		A value	
Organs	T. zillii	S. schall	p value	
Gills	5.33ª	5.29 ^a	p > 0.05	
Liver	5.75ª	5.37a	<i>p</i> > 0.05	
Muscles	5.94ª	5.28a	<i>p</i> > 0.05	
P value	p > 0.05	p > 0.05		

Note: across each row and down each column, similar superscripts indicate no significant difference (p > 0.05), while dissimilar superscripts indicate significant difference (p < 0.05).

DISCUSSION

Freshwater fish species have been described as wholesome foodstuffs, and its consumption is highly recommended by various nutritional health authorities (Egun, Oboh, 2022b; Egun et al., 2022; Egun et al., 2024). However, the ability of fishes to bioaccumulate toxic and non-biodegradable trace metals in various body parts

from the aquatic environment, has been raising serious concerns about the sustainable productivity of fishery resources and safety of fish food to consumers (Serviere-Zaragoza et al., 2021; Egun et al., 2023; Zaghloul et al., 2024). In this study, species variation in the accumulation of metals in the organs of T. zillii and S. schall was observed: the highest content of Fe, Zn, Mn and Pb was recorded in the liver and the highest content of Cu, Ni and Cr was found in the muscles of both fish species. The observed variations in the levels of accumulation of these metals in different organs of the fish species can be attributed to the differences in the physiological role of each organ and fish species. In similar studies, trace metal accumulation order of Fe > Zn > Cu > Pb > Cd was recorded in the gills, liver and muscles of Clarias gariepinus and T. zillii from the Owan River (Egun et al., 2025), while Fe > Zn > Mg > Cu > Pb > Cd (gills and muscles) and Fe > Zn > Cu > Mg > Pb > Cd (liver) in C. gariepinus and T. zillii from Ikpoba reservoir (Egun et al., 2023). That the observed similarity in the order of bioaccumulation of metals in this study compares with reported similar studies is indicative of the predisposition of these fish organs to readily take up iron, zinc, and copper from their environment. The unique structure of the gills for the direct absorption of trace metals from the environment and the role of the liver in biotransformation and detoxification makes these organs prominent sites for accumulation of metals. These factors including sex, fish habitat, and level of water pollution have been reported to influence the accumulation of metals in fish tissues (Nussey et al., 2000; Zaghloul et al., 2024).

Essential micronutrients – Fe, Zn, Cu, and Mn were present in good quantities in the fishes, which are attributed to their various functions as co-factors for the activation of a number of enzymes needed to regulate and maintain homeostatic status in fish (Kumar et al., 2020). The content of Fe, Zn, and Cu in the muscles of *T. zillii* and *S. schall* did not exceed their respective permissible concentrations of 100 mg/kg for Fe and 30 mg/kg for Zn and Cu each in fish

food (FAO/WHO, 2014). The content of Mn in the muscles of T. zillii (6.73 mg/kg) and S. schall (5.41 mg/kg) was very high and exceeded the WHO limit of 2.50 mg/kg in fish and fish products, although the high content of Fe, Zn, Cu, and Mn recorded in the muscles T. zillii and S. schall makes them good sources of mineral elements to consumers. Several toxicological studies reported negative effects of the bioaccumulation of these metals on the physiology of fishes exposed to elevated levels of these metals in their habitats (Akan et al., 2012; Simionov et al., 2019; Zeeshan et al., 2021; Zaghloul et al., 2024). Similar studies have reported high Fe and Zn contents in the muscles of T. zillii from Ikpoba reservoir (Egun, Oboh, 2022) and the Owan River (Egun et al., 2022b); lower Cu (0.25-0.52 mg/kg) and Mn (0.52 mg/kg) contents in T. zillii from the Ogbese River (Olayinka-Olagunju, 2021), and a higher content of Mn was recorded in the muscles of T. zillii from Ogbainbiri (22.67 mg/Kg) and Samabiri (14.64 mg/Kg) along the Nun River in Bayelsa State (Aigberua, Tarawou, 2017).

The presence of non-essential toxic trace elements - Ni, Cr, and Pb - in the organs of the fish species, especially in fish muscles, is an indication of fish meat contamination. The content of Ni and Cr in the muscles of T. zillii and S. schall exceeded the maximum permissible limit of 0.4 mg/kg for Ni in fish meat (EU Commission Regulation, 2006), and WHO/FEPA maximum allowable limit for Cr (0.05-0.15 mg/kg) in fish food (FEPA, 2003; Bakshi, Panigrahi, 2018). For Pb, its content in the muscles of T. zillii and S. schall was very low and below the maximum permissible limit of 0.3 mg/kg for lead in fish meat (FAO/ WHO, 2010). These non-essential toxic trace elements have been reported to cause several disruptive effects on the growth and development of fishes even at very low concentrations (Burger et al., 2002; Krishna et al., 2014). Also, the carcinogenic potentials of Ni, Cr, and Pb portends a concern for consumers and public health, as human exposure to these metals has been linked to several ailments in adults and children (Umar et al., 2021)

Metal Pollution Index (MPI)

The Metal Pollution Index (MPI) indicates the degree of bioaccumulation of metals within a biota and has garnered interest as a valuable tool for estimating metal contamination (USEPA, 2011). In this study, high values of the MPI, particularly in fish muscles, is a cause of concern for food safety and public health, as it is an indication of the degree of metal pollution of the Siluko River and high susceptibility of these metals to bioaccumulate in the fishes. Variations in metal accumulation and distribution in fish organs have been attributed to the route of organ exposure to the metal source such as through diet or by direct contact with polluted water (Zaghloul et al., 2024).

CONCLUSIONS

This study assessed the concentrations of trace metals in T. zillii and S. schall from the Siluko River. The identified metals - Fe, Zn, Cu, Mn, Ni, Cr, and Pb - were detected in substantial amounts. The high content of essential trace elements - Fe, Zn, Cu, and Mn - recorded in the muscles T. zillii and S. schall makes them good sources of the mineral elements to consumers. However, the elevated levels of these essential trace elements and the non-essential toxic elements - Ni, Cr and Pb - in the fishes as indicated by the MPI values predisposes the physiology of the fishes to metal toxicity and portends health risk to consumers. There is a need for (1) the identification of the sources of these heavy metal pollutants with the Siluko River catchment area, (2) commencement of a management strategy that will ensure a decline in the inflow of pollutants into the river, and (3) continuous monitoring of heavy metal pollution levels in water and fish to guarantee public health.

ACKNOWLEDGEMENTS

The authors are grateful to Mr Eseoghene Erhurhu, a postgraduate student at the Department of Animal and Environmental Biology, University of Benin, for his assistance in the collection,

identification, and preparation of fish samples, also to Dr Peter Uwaifo, Mr Timothy Agho, and all the laboratory technicians at Benin Owena River Basin/University of Benin Joint Analytical Laboratory that have contributed to producing high-quality analytical data.

DECLARATIONS

The authors have no relevant financial or non-financial interests to disclose.

The authors have no competing interests to declare that are relevant to the contents of this article.

Received 8 April 2025 Accepted 13 May 2025

References

- 1. Aigberua A, Tarawou T. Assessment of Heavy Metals in Muscle of Tilapia zilli from Some Nun River Estuaries in the Niger Delta Region of Nigeria. J Sci. 2017;2(9):96–101.
- 2. Akan JC, Salwa M, Bashir SY, Victor OO. Bioaccumulation of some heavy metals in fish samples from River Benue in Vinikilang, Adamawa State, Nigeria. Amer J Anal Chem. 2012;3:727–36.
- 3. APHA/AWWA/WEF. Standard methods for the examination of water and wastewater. Washington, DC: American Public Health Association; 2012.
- 4. Bakshi A, Panigrahi AK. A comprehensive review on chromium induced alterations in fresh water fishes. Toxicol Rep. 2018;5:440–7. https://doi.org/10.1016/j.toxrep.2018.03.007
- 5. Basooma A, Teunen L, Semwanga N, Bervoets L. Trace metal concentrations in the abiotic and biotic components of River Rwizi ecosystem in western Uganda, and the risks to human health. Heliyon. 2021;7(11):e08327. https://doi.org/10.1016/j.heliyon.2021
- 6. Bawuro AA, Voegborlo RB, Adimado AA. Bioaccumulation of heavy metals in some tis-

- sues of fish in Lake Geriyo, Adamawa State, Nigeria. J Environ Pub Health. 2018;1854892. https://doi.org/10.1155/2018/1854892
- Biose E, Egun NK, Uzoh N, Omoigberale MO. Assessment of water quality and associated human health risk of a tropical freshwater body in Edo State, Nigeria. Afr J Aquat Sci. 2024, 1–12. https://doi.org/10.2989/1608591 4.2023.2292125
- 8. Burger J, Gaines KF, Boring CS, Stephens WL, Snodgrass J, Dixon C, McMahon M, Shukla S, Shukla T, Gochfeld M. Metals levels in fish from the Savannah River: potential hazards to fish and other receptors. Environ Res. 2002;89(1):85–97.
- Chen L, Wang J, Beiyuan J, Guo X, Wu H, Fang L. Environmental and health risk assessment of potentially toxic trace elements in soils near uranium (U) mines: a global meta-analysis. Sci Total Environ. 2022;816:151556. https:// doi.org/10.1016/j.scitotenv.2021.151556
- Custer TW, Custer CM, Hines RK, Sparks DW. Trace elements, organochlorines, polycyclic aromatic hydrocarbons, dioxins, and furans in lesser scaup wintering on the Indiana Harbor Canal. Environ Pollut. 2000;110:469–82. https://doi.org/10.1016/s0269-7491(99)00315-2
- 11. Egun NK, Oboh IP, Olowo CU, Enabulele CO. Fish food and community health: an assessment of the contribution of capture fisheries to nutritional needs of households in Nigeria. Aquatic Food Studies. 2024;4(2):AFS260. https://doi.org/10.4194/AFS260
- 12. Egun NK, Oboh IP. Assessment of water quality for suitability and human health risk: a study of the Owan River, Edo State, Nigeria. Afr J Aquat Sci. 2023. https://doi.org/10.2989/16085914.2022.2156468
- 13. Egun NK, Oboh IP. Surface water quality evaluation of Ikpoba reservoir, Edo State, Nigeria. IJEWR. 2022a;6:509–19. https://doi.org/10.1007/s42108-021-00139-z
- 14. Egun NK, Oboh IP. Potential contribution of captured fishes to the recommended nutrient intakes (RNIs): a case study of commercial

- fish species from Ikpoba reservoir, Edo State, Nigeria. Measurement: Food. 2022b;5. https://doi.org/10.1016/j.meafoo.2021.100014
- Egun NK, Ogiesoba-Eguakun CU. Physico – Chemical and Water Quality Index Analysis of the Okhuaihe River, Edo State Nigeria. Afr J Aquat Sci. 2018;43:(4):345–51.
- 16. Egun NK, Okotie UR, Oboh IP. Distribution and toxicological risk assessment of metals in commercial fish species from a lentic ecosystem in Nigeria. J Trace Elem Min. 2023;6:100100. https://doi.org/10.1016/j. jtemin.2023.100100
- 17. Egun NK, Olowo UC, Udeh H, Oboh IP. Toxicological assessment of captured fish species from a lotic freshwater ecosystem in Nigeria. Ecotoxicol Environ Contam. 2025;19(2):31–40. https://doi.org/10.5132/eec.2024.02.04
- 18. Egun NK. Effect of water quality on the nutritional composition of *Clarias gariepinus* and *Tilapia zillii* from Ikpoba Reservoir and Owan River, Edo State, Nigeria. PhD thesis, 2021. University of Benin, Nigeria.
- 19. Egun NK, Olowo UC, Oboh IP. Potential contribution of selected wild fish species to the minerals intake of pregnant and lactating women, children and adults in rural riverine communities of Edo State: insights and outcomes. Measurement: Food. 2022;8. https://doi.org/10.1016/j.meafoo.2022.100063
- 20. Enuneku A, Ezemonye LI, Adibeli F. Heavy metal concentrations in surface water and bioaccumulation in fish (*Clarias gariepinus*) of River Owan, Edo State, Nigeria. EIJST. 2013;2(7):31–9.
- 21. Commission Regulation (EC) No. 1881/2006 of 19 December 2006 setting maximum levels for certain contaminants in foodstuffs. 2006. Available online: https://www.legislation.gov.uk/eur/2006/1881 [cited 15 September 2022]
- 22. FAO/WHO. FAO Fisheries and Aquaculture Report No. 978; Proceedings of the Joint FAO/WHO Expert Consultation on the Risks and Benefits of Fish Consumption; Rome, Italy. 25–29 January 2010.

- 23. FAO/WHO (2011). Joint FAO/WHO food standards programme codex committee on contaminants in foods. 5th Session, The Hague Netherlands, 21–25 March, 2011, http://ftp.fao.org/codex/meetings/CCCF/cccf5/cf05_INF.pdf
- Federal Environmental Protection Agency (FEPA). Guidelines and standards for environmental pollution control in Nigeria. 2003; 238.
- Idodo-Umeh G. Freshwater fishes of Nigeria (taxonomy, ecological notes, diet and utilization). Benin City: Idodo Umeh Publishers Limited, 2003.
- 26. Islam MA, Das B, Quraishi SB, Khan R, Naher K, Hossain SM, Karmaker S, Latif SA, Hossen MB. Heavy metal contamination and ecological risk assessment in water and sediments of the Halda river, Bangladesh: a natural fish breeding ground. Mar Pollut Bull. 2020;160:111649. https://doi.org/10.1016/j.marpolbul.2020.111649
- 27. Juśkiewicz W, Gierszewski P. Toxic metal pollution of aquatic ecosystems of European Union nature protection areas in a region of intensive agriculture (Lake Gopło, Poland). Aquat Sci. 2002;84:52. https://doi.org/10.1007/ s00027-022-00884-1
- 28. Krishna PV, Rao KM, Swaruparani V, Rao DS. Heavy metals concentration in fish *Mugil cephalus* from Machilipatnam coast and possible health risks to fish consumers. Br Biotechnol J. 2014;4(2):126–35.
- 29. Kumar N, Krishnani KK, Singh NP. Effect of zinc on growth performance and cellular metabolic stress of fish exposed to multiple stresses. Fish Physiol Biochem. 2020;46:315–29.
- 30. Muhammad S, Ahmad K. Heavy metal contamination in water and fish of the Hunza River and its tributaries in Gilgit–Baltistan: evaluation of potential risks and provenance. Environ Technol Innov. 2020;20:101159. https://doi.org/10.1016/j.eti.2020.101159
- 31. Muneer J, AlObaid A, Ullah R, Ur Rehman K, Erinle KO. Appraisal of toxic metals in water, bottom sediments and fish of fresh water lake.

- J King Saud Univ Sci. 2022;34:101685. https://doi.org/10.1016/j.jksus.2021.101685
- 32. Nussey G, Van Vuren JHJ, Du-Preez HH. Bio-accumulation of chromium, manganese, nickel and lead in the tissues of the moggel, *Labeo umbratus* (Cyprinidae), from Witbank Dam, Mpumalanga. Water SA. 2000;26(2):269–84.
- 33. Oboh IP, Agbala CS. Assessment of the quality of River Siluko, Edo State, Southern Nigeria using water quality index (WQI). Afr J Aquat Sci. 2017;42(3):279–86.
- 34. Olayinka-Olagunju JO, Dosumu AA, Olatunji-Ojo AM. Bioaccumulation of heavy metals in pelagic and benthic fishes of Ogbese River, Ondo State, South-Western Nigeria. Water Air Soil Pollut. 2021;232(44). https://doi.org/10.1007/s11270-021-04987-7
- 35. Qadir A, Malik RN. Heavy metals in eight edible fish species from two polluted tributaries (Aik and Palkhu) of the River Chenab, Pakistan. Biol Trace Elem Res. 2011;143(3):1524–40.
- 36. Rainbow PS. Trace metal bioaccumulation: models, metabolic availability and toxicity. Environ Int. 2007;33(4):576–82. https://doi.org/10.1016/j.envint.2006.05.007
- 37. Serviere-Zaragoza E, Lluch-Cota SE, Mazariegos-Villarreal A, Balart EF, Valencia-Valdez H, Méndez-Rodríguez LC. Cadmium, lead, copper, zinc, and iron concentration patterns in three marine fish species from two different mining sites inside the Gulf of California, Mexico. Int J Environ Res Public Health. 2021;18(2):844.
- 38. Simionov IA, Cristea V, Petrea SM, Mogodan A, Nicoara M, Baltag ES, Strungaru SA, Faggio C. Bioconcentration of essential and nonessential elements in black sea turbot (Psetta maxima Maeotica Linnaeus, 1758) in relation to fish gender. J Mar Sci Eng. 2019;7(12):466.
- 39. Standard Reference Material (SRM) 3233. National Institute of Standards and Technology Certificate of Analysis. 2020. https://tsapps.nist.gov/srmext/certificates/3233.pdf

- 40. Tesfaye T, Tesfau H, Abebe D. et al. Assessment of trace metal levels in water, sediment and fish tissue from Lake Small Abaya, Ethiopia. Discov Environ. 2024;2:124. https://doi.org/10.1007/s44274-024-00159-6
- 41. Umar A, Umar R, Ahmad MS. Hydrogeological and hydrochemical frame works of regional aquifer system in Kali-Ganga sub-basin. India. Environ Geol. 2021;40(4–5):602–11.
- 42. US Environmental Protection Agency (USEPA). Regional screening level (RSL) summary table: November 2011. United States Environmental Protection Agency, Washington, DC, Philadelphia. http://www.epa.gov/regshwmd/risk/human/Index.htm
- 43. Uwaifo OP, Egun NK, Agho TI. Ecotoxicological risk assessment of metalloid contamination in the sediment and benthic fauna of a tropical lotic freshwater ecosystem in Southern Nigeria. Biologija. 2023;69(3):217–31. https://doi.org/10.6001/biologija.2023.69.3.2
- 44. Zaghloul GY, Eissa HA, Zaghloul AY et al. Impact of some heavy metal accumulation in different organs on fish quality from Bardawil Lake and human health risks assessment. Geochem Trans. 2024;25:1. https://doi.org/10.1186/s12932-023-00084-2
- 45. Zahoor I, Mushtaq A. Water pollution from agricultural activities: a critical global review. Int J Chem Biochem Sci. 2023;23:164–76.
- 46. Zeeshan A, Ali MY, Nadia S, Ijaz M, Gul E, Nayab SAM, Syed TS, Michael A, Ijaz K, Haroon K. Toxicity and bioaccumulation of manganese and chromium in different organs of common carp (*Cyprinus carpio*) fish. Toxicol Rep. 2021;8:343–8. https://doi.org/10.1016/j. toxrep.2021.02.003

Egun Nkonyeasua Kingsley, Olowo Ukpamufo Cyril, Biose Ekene, Jacob N. Jacob, Oboh Ijeoma Patience

MIKROELEMENTŲ PASISKIRSTYMO SILUKO UPĖS (EDO VALSTIJA, NIGERIJA) *TILAPIA* ZILLII IR SYNODONTIS SCHALL ŽUVYSE TYRIMAS

Santrauka

Sunkiųjų metalų tarša vandens ekosistemose kelia susirūpinimą dėl jų kaupimosi vandens organizmuose ir tolesnio biomagnifikacijos poveikio vandens mitybos grandinėje. Šiuo tyrimu buvo nustatytas mikroelementų pasiskirstymas Siluko upės Tilapia zillii ir Synodontis schall žuvyse Edo valstijoje, Nigerijoje. Švieži žuvų mėginiai buvo renkami kas mėnesį nuo 2021 m. rugsėjo iki 2022 m. rugpjūčio mėnesio kartu su vietiniais žvejais. Mikroelementų kiekiai žiaunose, kepenyse ir raumenyse buvo nustatyti atomų absorbcinės spektroskopijos metodu. Metalų taršos indeksu (MPI) buvo įvertintas žuvų metalų bioakumuliacijos lygis. Rezultatai atskleidė tarprūšinius metalų kaupimosi įvairiuose organuose skirtumus: T. zillii turėjo didesnį vidutinį vario (Cu), cinko (Zn), mangano (Mn) ir geležies (Fe) kieki visuose tirtuose organuose, o S. schall - didesnį nikelio (Ni), chromo (Cr) ir švino (Pb) kiekį. MPI reikšmės svyravo nuo 5,28 iki 5,94 T. zillii ir S. schall organuose. T. zillii atveju MPI reikšmės mažėjo tokia tvarka: raumenys > kepenys > žiaunos, o S. schall – kepenys > žiaunos > raumenys. Gauti rezultatai patvirtina su vandens tarša susijusį žuvų užterštumą mikroelementais. Didelės MPI reikšmės rodo Siluko upės taršos metalais lygį ir ryškią jų bioakumuliacijos žuvų audiniuose tendenciją, ir tai kelia pavojų žmonių sveikatai vartojant žuvis maistui. Rekomenduojama nuolatinė veiklos stebėsena Siluko upės baseine siekiant sumažinti sunkiųjų metalų taršą.

Reikšminiai žodžiai: toksiškumas vandens organizmams, bioakumuliacija, metalų toksiškumas, metalų taršos indeksas (MPI), Siluko upė